

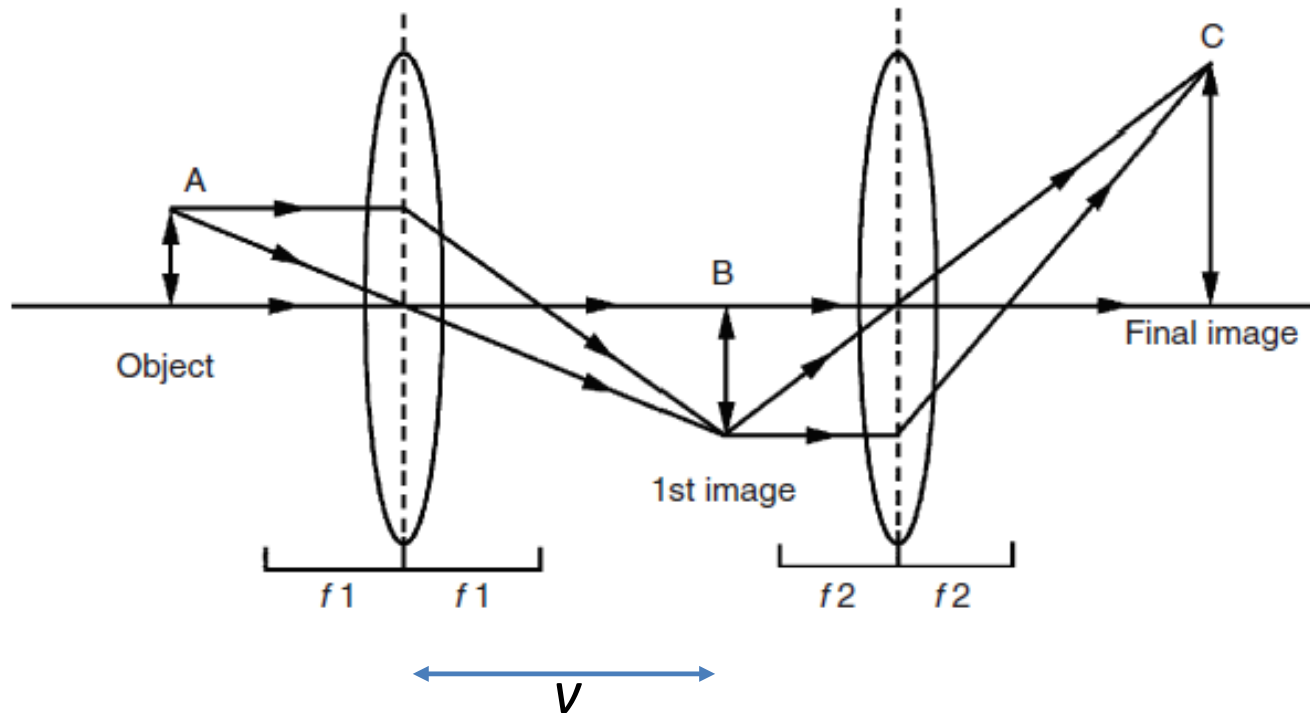
Optical Microscopy: structure and working principles

Lecturer: Kung-Hsuan Lin

TIGP Course: Characterization, Fabrication,
and Manipulation at Nanometer Scale

March 26, 2020

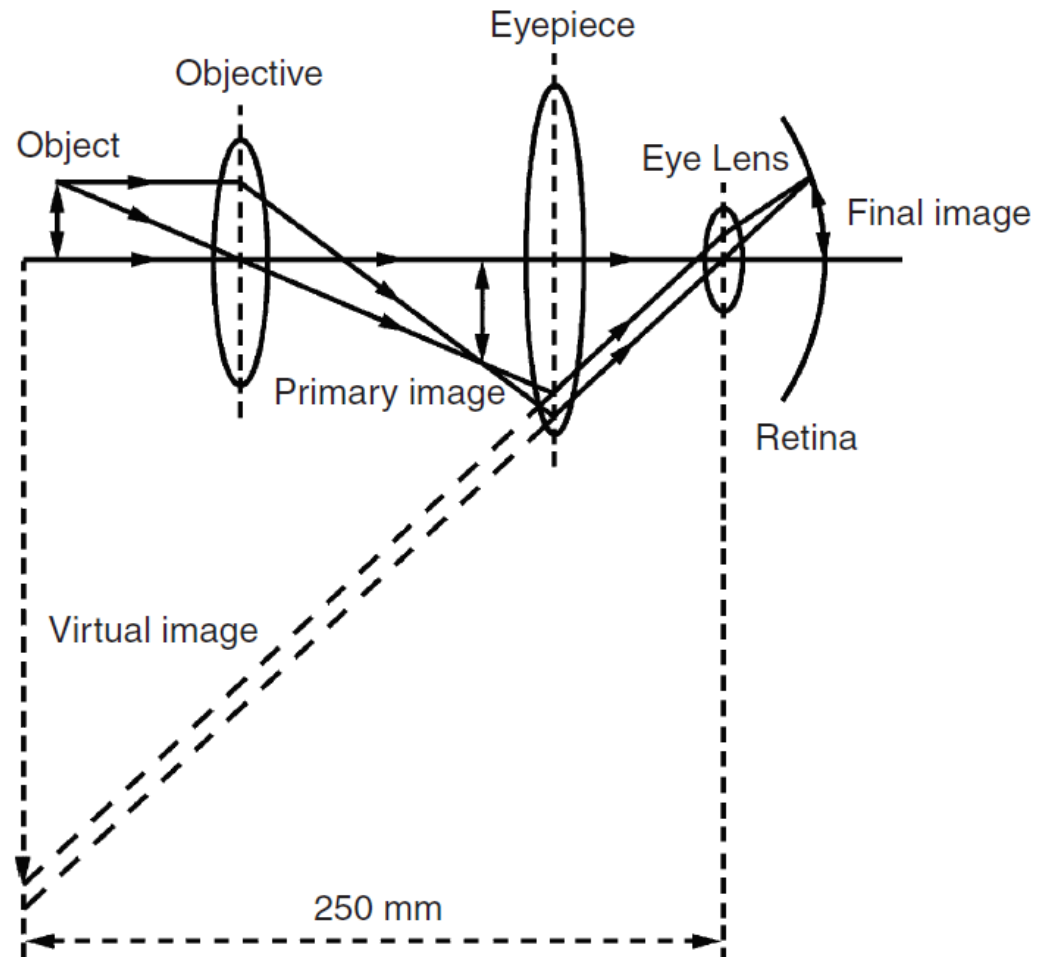
Image Formation



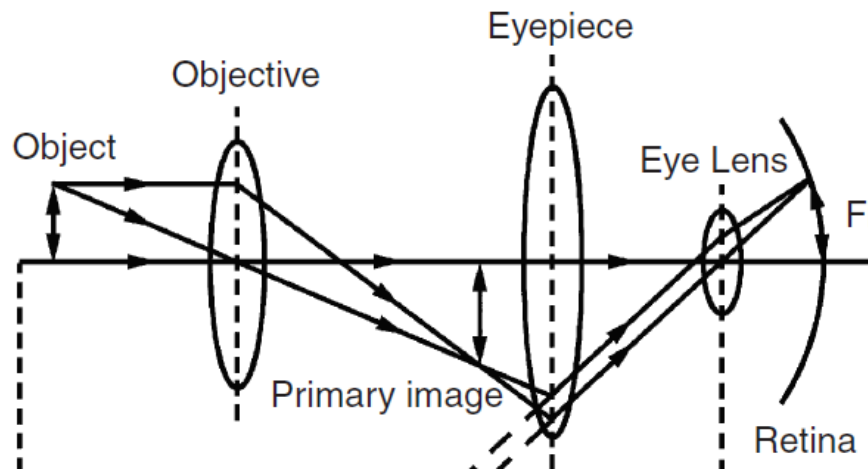
$$M = \frac{v - f}{f}$$

$$M = M_1 M_2 \frac{(v_1 - f_1)(v_2 - f_2)}{f_1 f_2}$$

Image observed by eyes



Tube length & Infinity corrected optics



↔
Tube length:
standardized to the Royal Microscopical
Society (RMS) suggestion of 160 mm

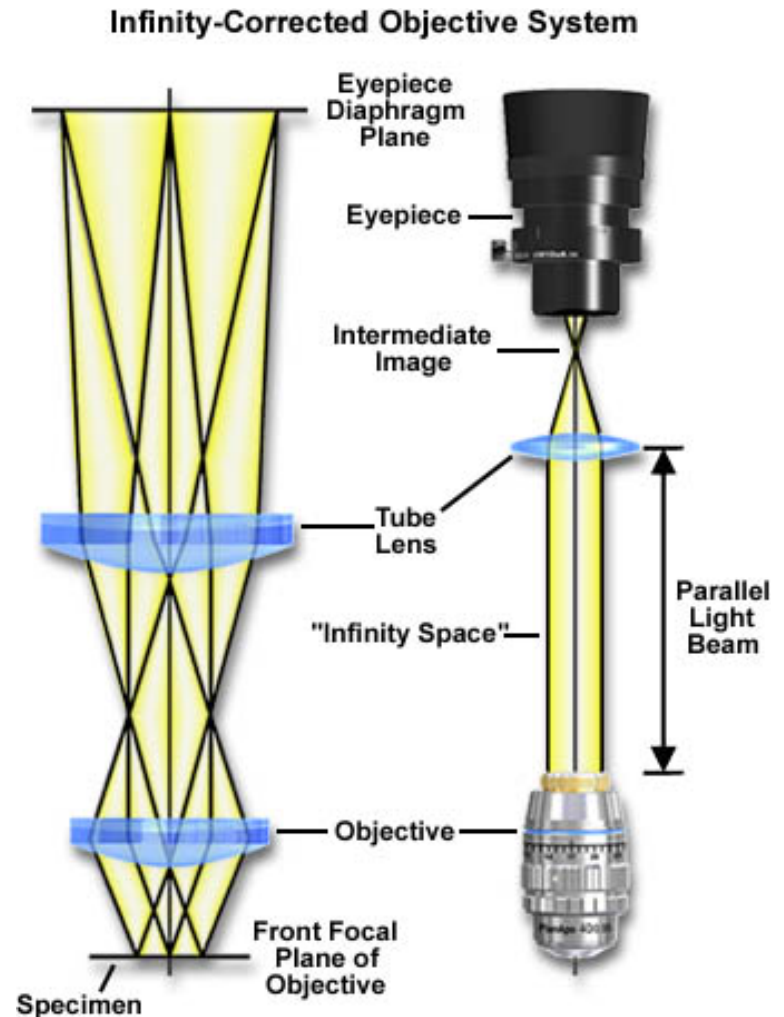


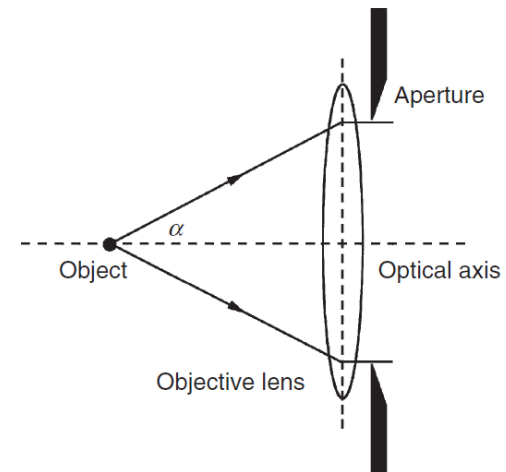
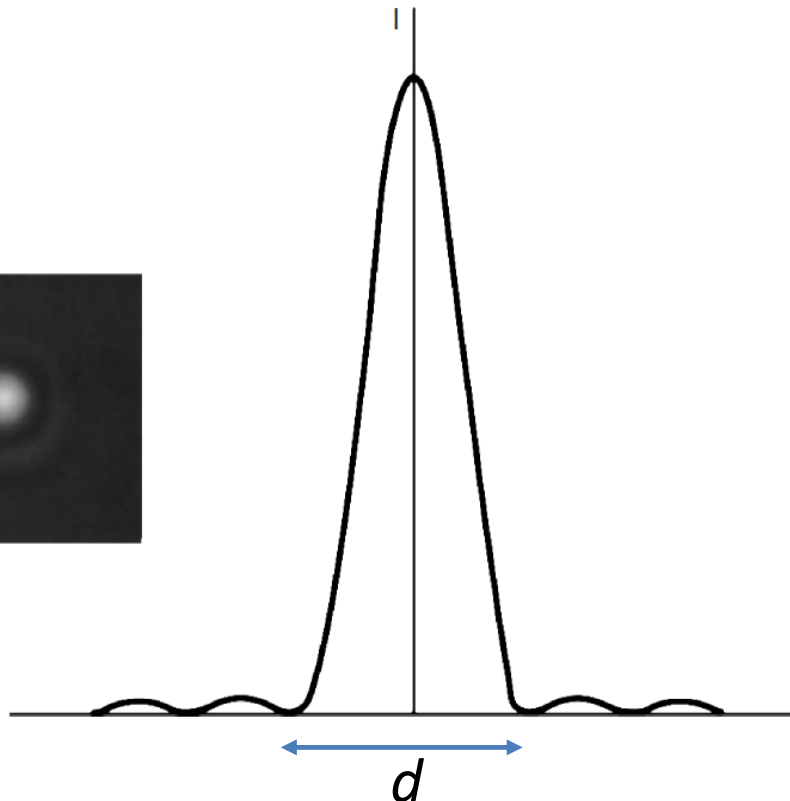
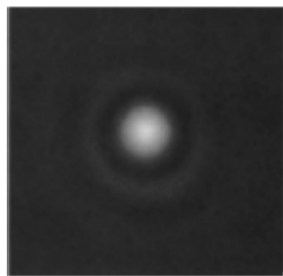
Figure 3

Diffraction limitation

- Due to diffraction, the smallest point to which a can focus a beam of light is the size of the **Airy disk**. The smallest size is **diffraction limited**.

$$d = \frac{1.22\lambda}{n \sin \alpha} = \frac{1.22\lambda}{NA}$$

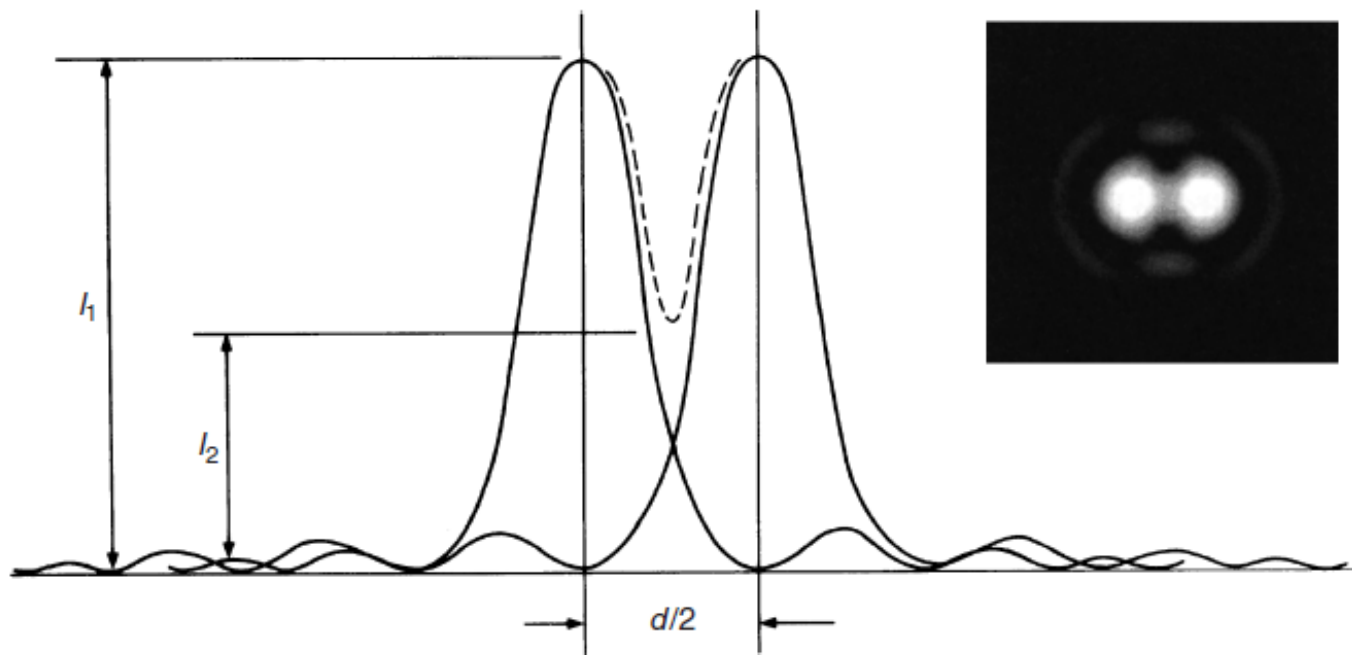
NA: numerical aperture



Optical resolution

- Resolution: the minimum distance to distinguish two point objects.

$$R \equiv d / 2 = \frac{0.61\lambda}{NA}$$



Effective magnification

$$R = d / \lambda = \frac{0.61\lambda}{NA}$$

Optical resolution: minimum ~ 200 nm for visible light

Eye resolution: minimum ~ 0.2 mm

(A microscope should enlarge features to the resolution level of the human eye



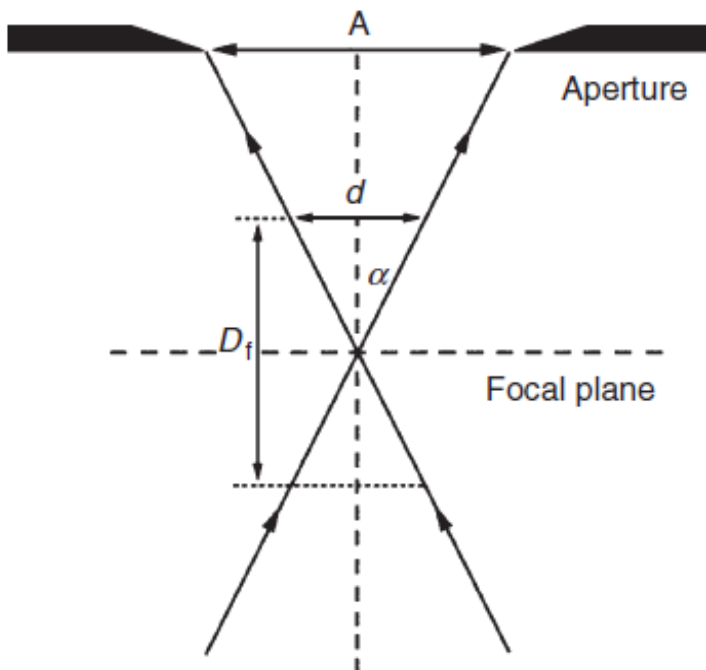
X



= 1000

Depth of Field

- The range of position for an object in which image sharpness does not change.



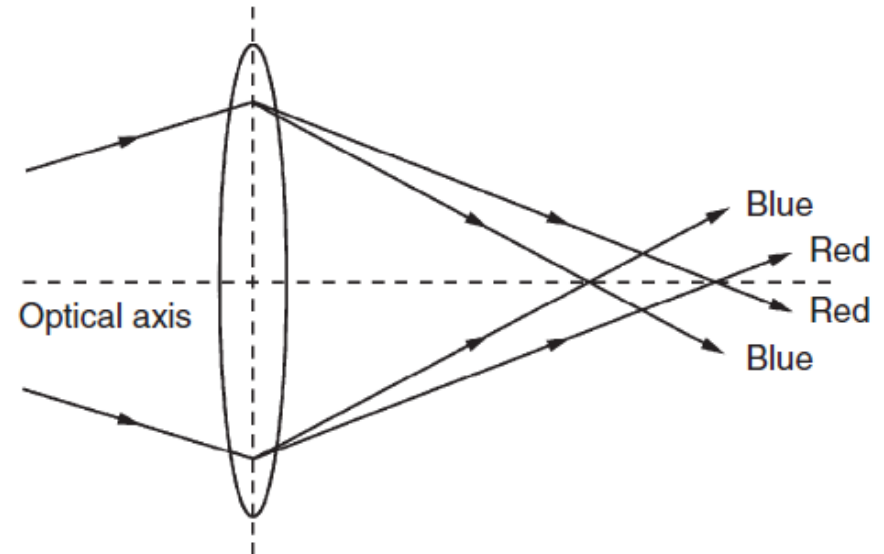
$$D_f \equiv \frac{d}{\tan \alpha}, \quad d = \frac{1.22\lambda}{NA}$$

Depth of focus

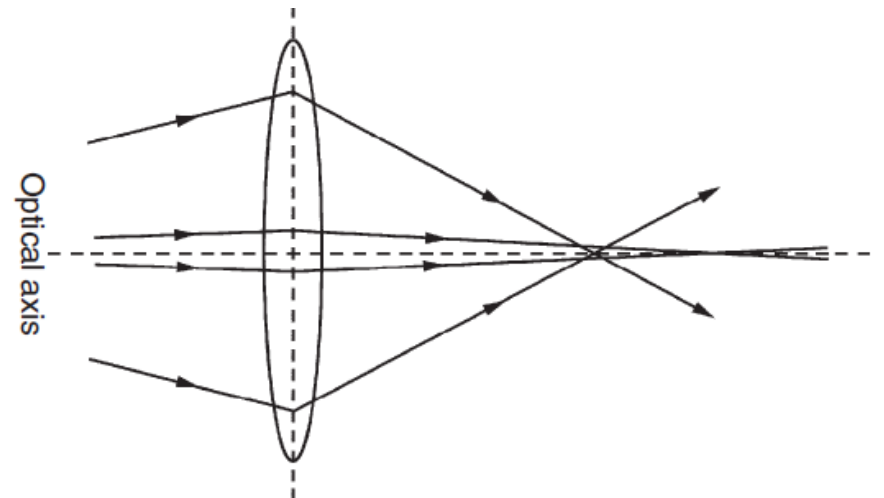
The range of image plane position which is viewed in focus.

Aberration

- Chromatic Aberration

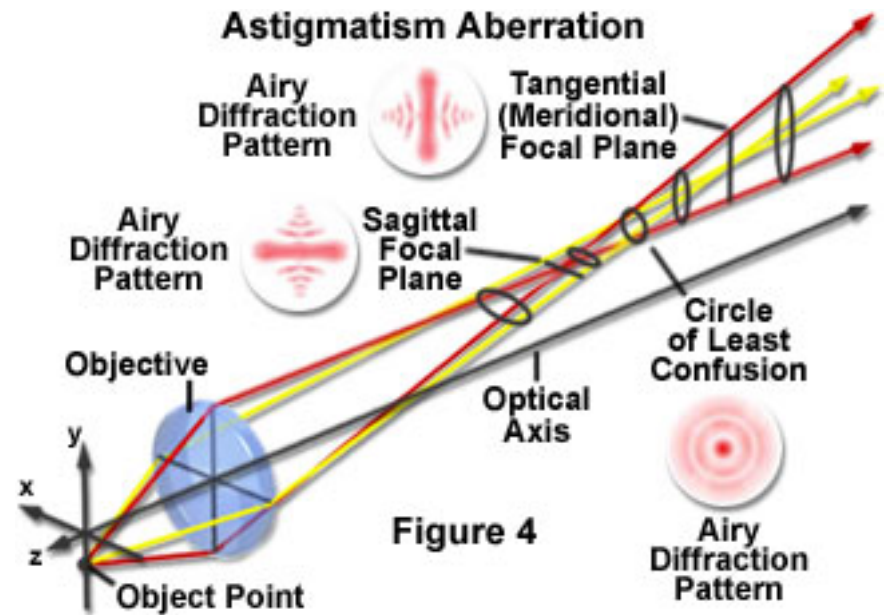


- Spherical Aberration

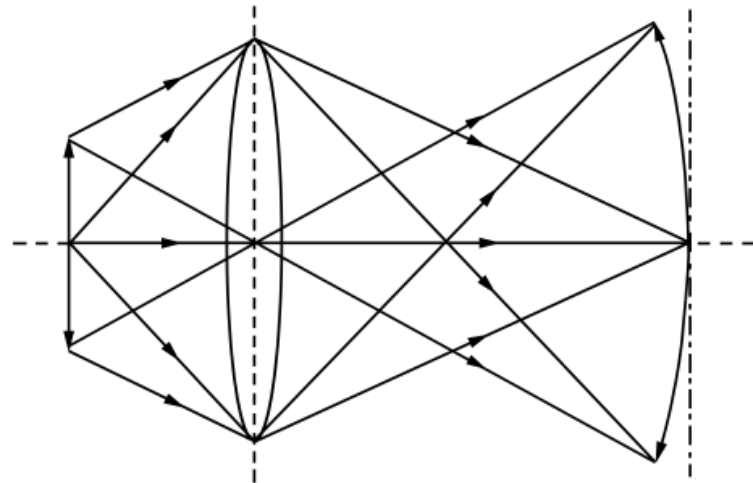


Aberration

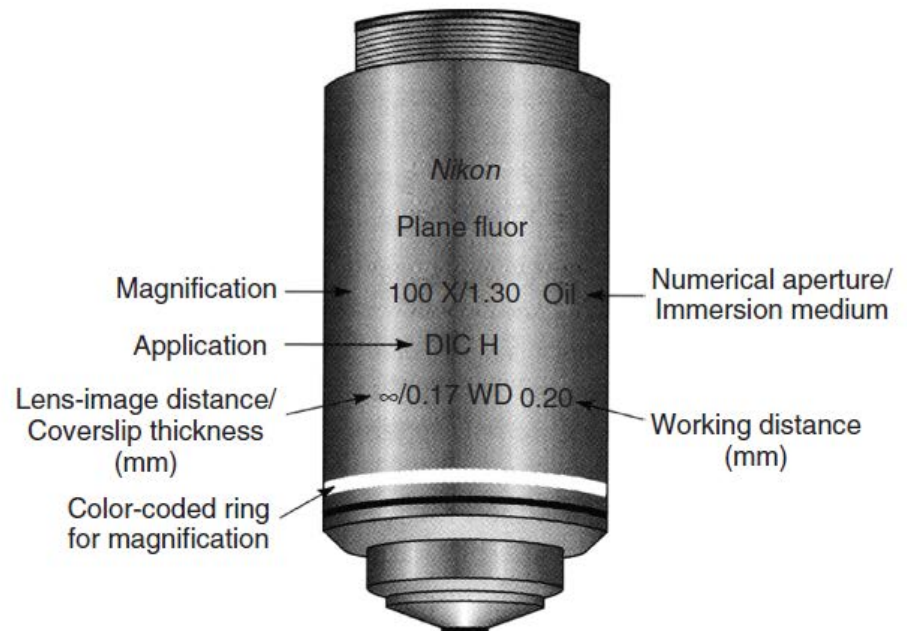
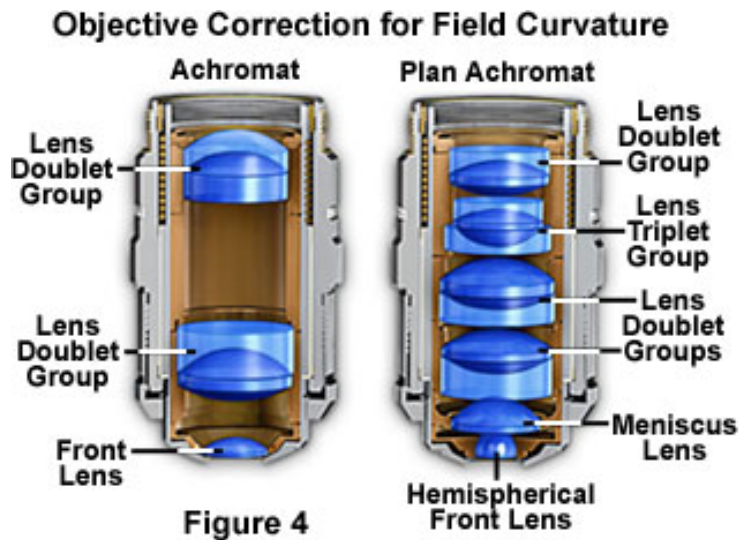
- Astigmatism Aberration



- Field Curvature Aberration



Objective lens specification

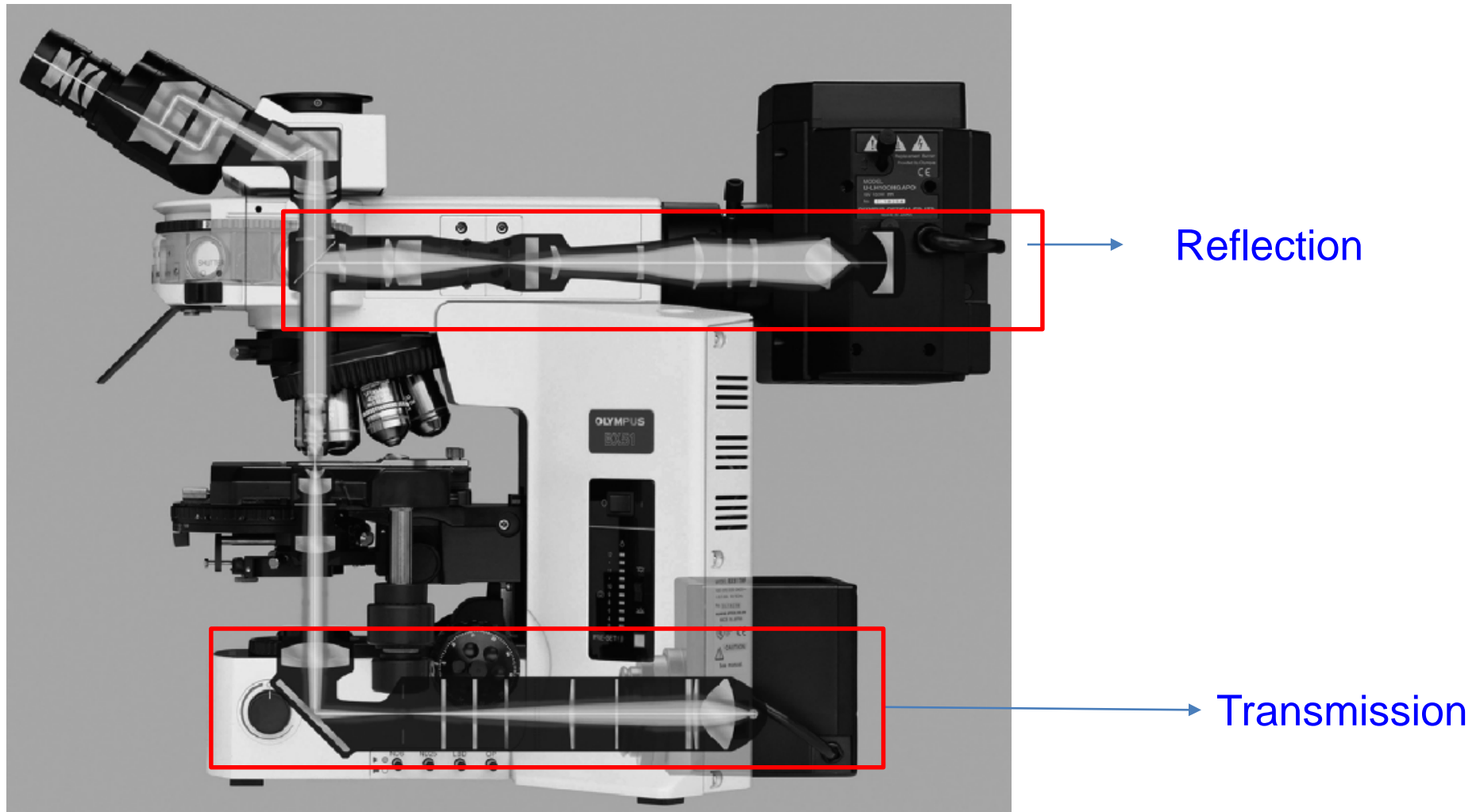


Achromat: correct red and blue

Semiachromat (fluorite): improvement of achromatic aberration

Apochromat: completely eliminate achromatic aberration. And spherical correction of two colors.

Köhler Illumination



Illumination light sources

- Incandescent Lamps

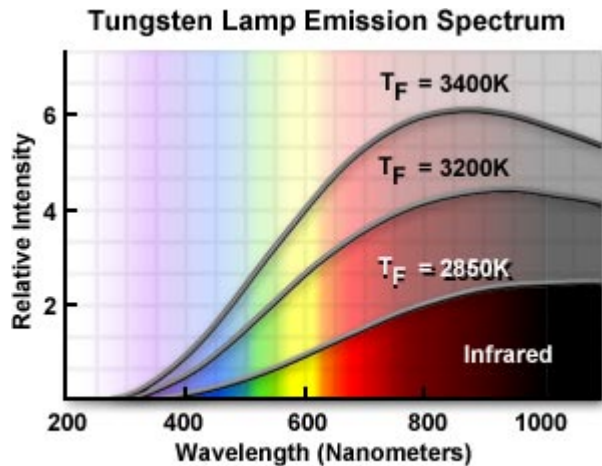


Figure 2

- Arc Lamps

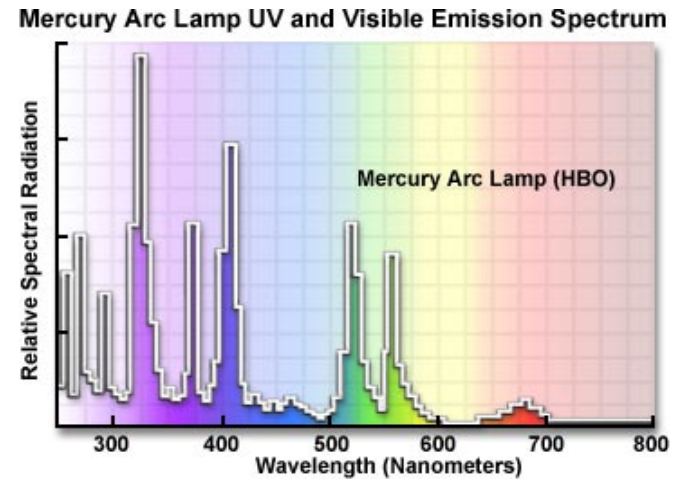
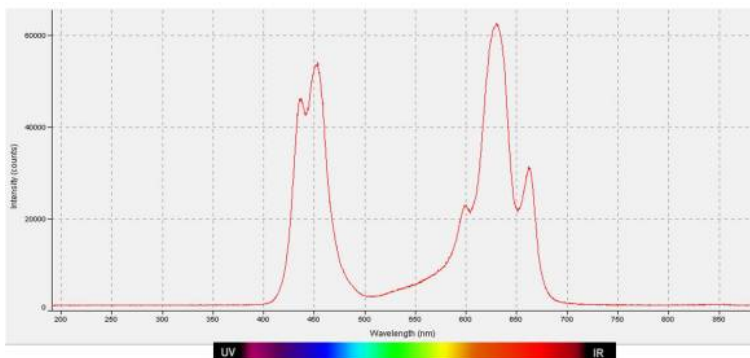


Figure 8

- LED



Xenon Arc Lamp Emission Spectrum

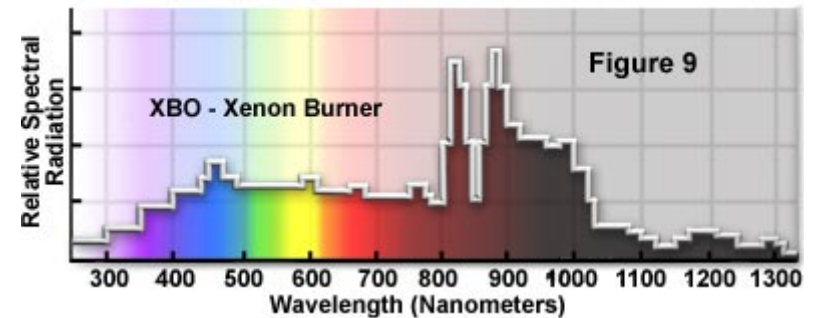
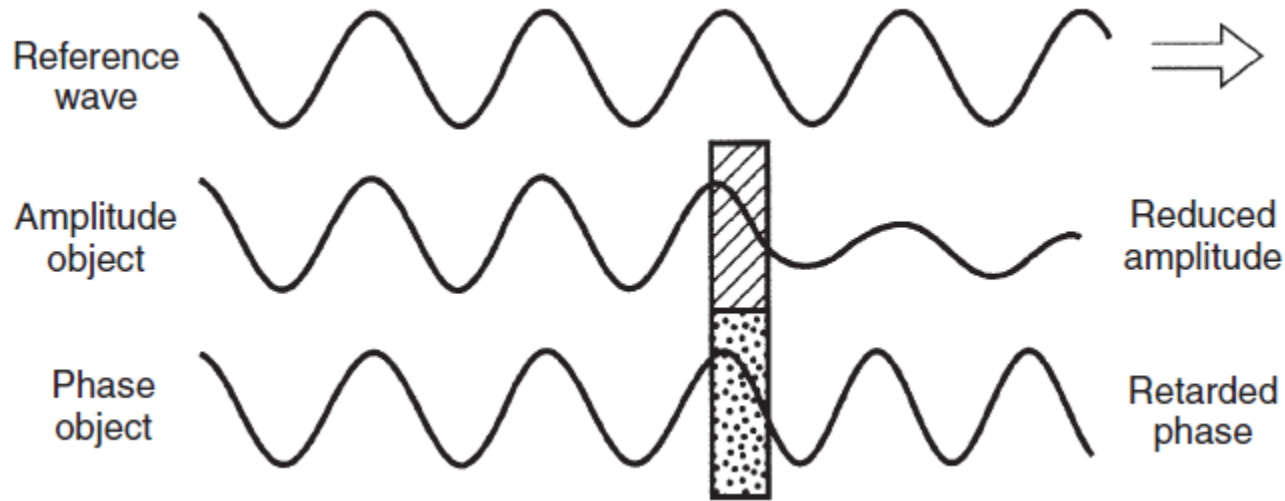


Figure 9

Imaging modes



Amplitude object
(visible)



Bright-field imaging
Dark-field imaging

Phase object
(invisible -> should be
converted to intensity)



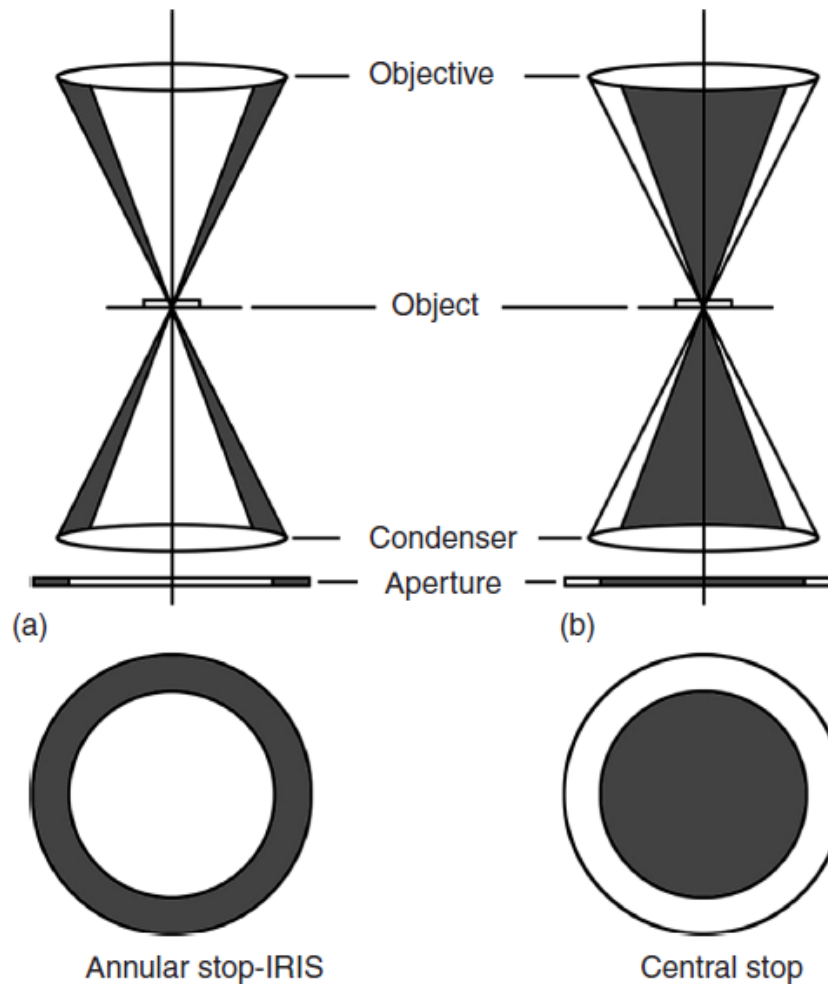
Phase contrast imaging
Differential interference contrast
(DIC) imaging

Fluorescence object
(visible)



Fluorescence imaging

Bright-Field and Dark-Field Imaging



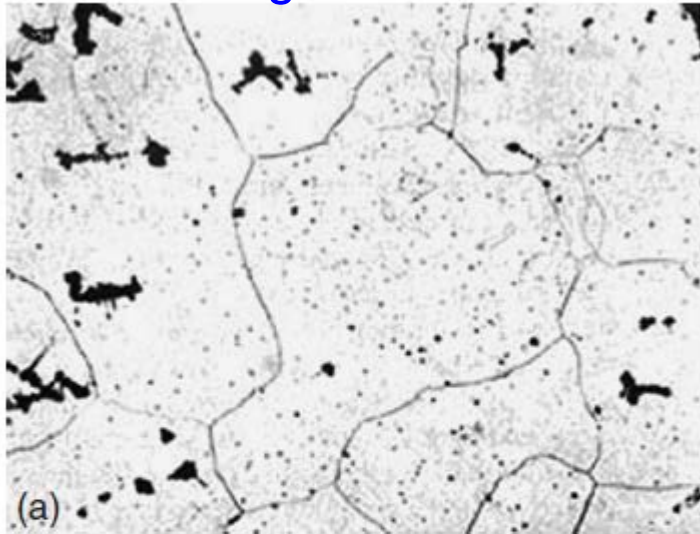
Bright-field illumination

Dark-field illumination

Contrast

$$\text{Contrast} = \frac{I_{\text{object}} - I_{\text{background}}}{I_{\text{background}}}$$

Bright-field



Dark-field

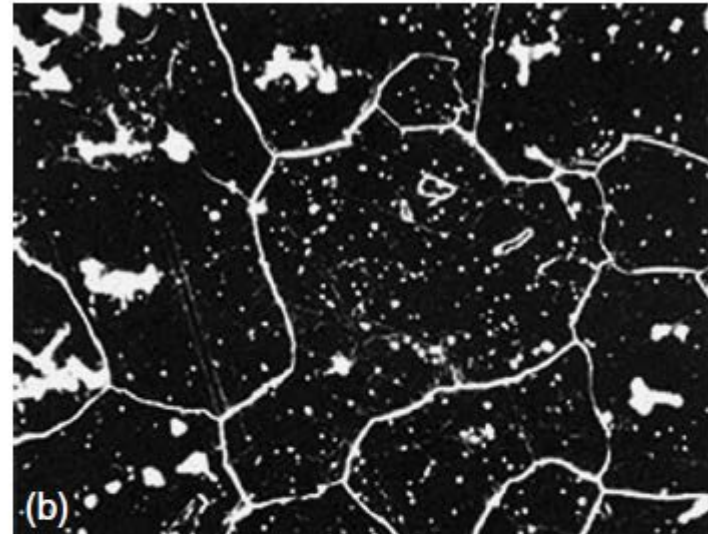
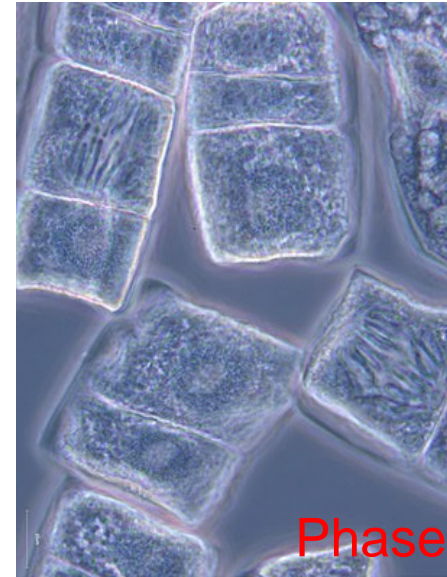
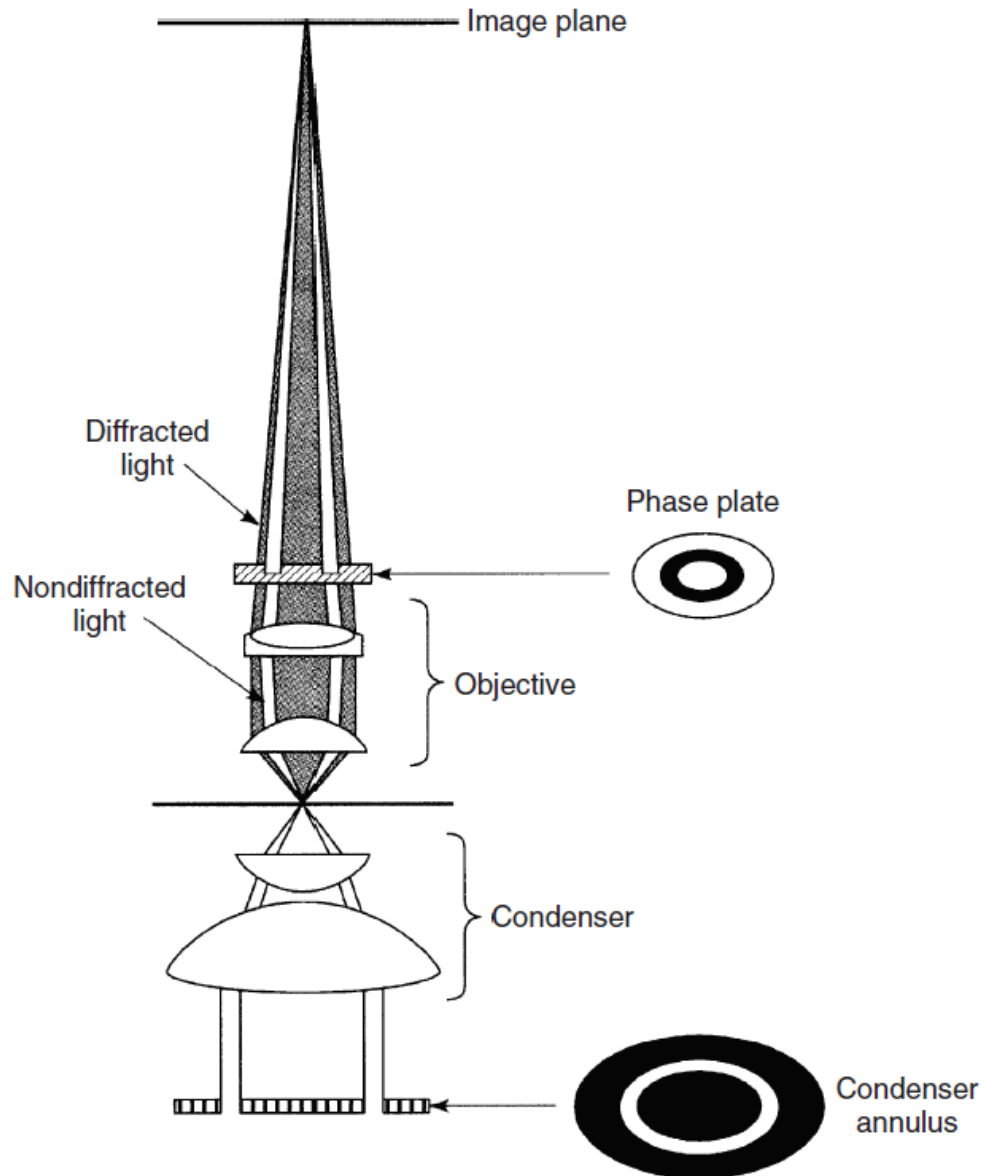


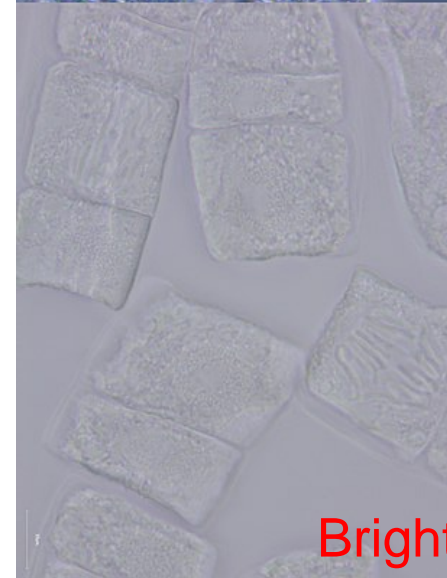
Figure 1.30 Comparison between: (a) bright-field and (b) dark-field images of AISI 1080 high carbon steel. In addition to grain boundaries and oxide particles, annealing twins are revealed in the dark-field

image. (Reproduced with permission of ASM International®. All Rights Reserved. www.asminternational.org. Ref. [2]. © 1984 ASM International®.)

Phase Contrast Microscopy



Phase Contrast

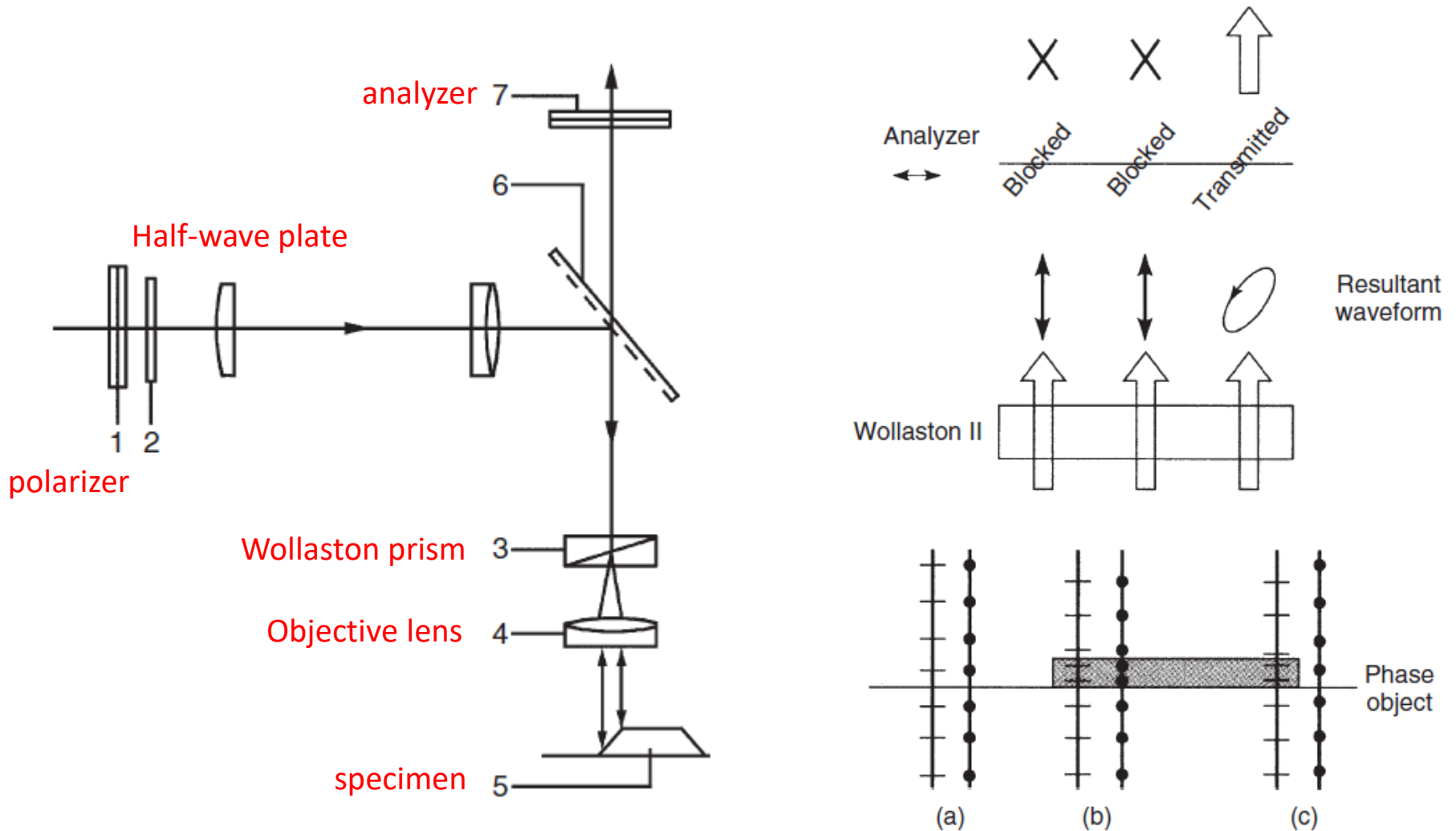


Bright-field

Onion Root Tip Cells

Nomarski Microscopy

Also called **Differential Interference Contrast (DIC)**



Comparison between phase contrast and DIC

DIC

(more stereo in height)

Phase Contrast

(high contrast, but artifact of halos)

Transparent Specimens in Phase Contrast and DIC

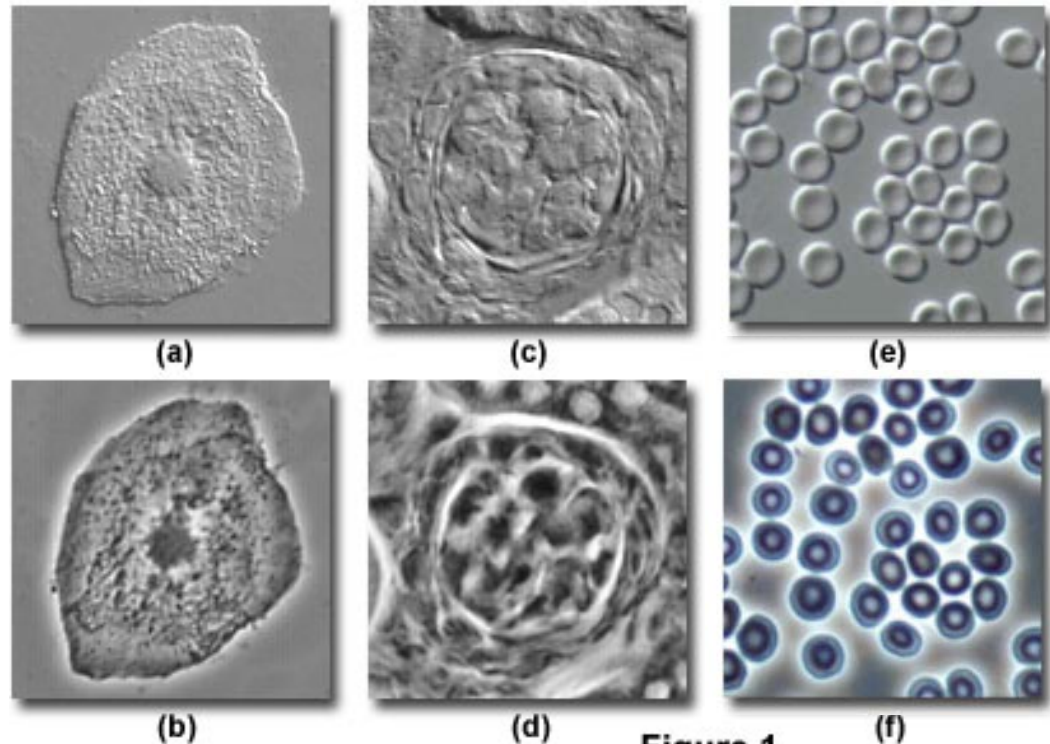
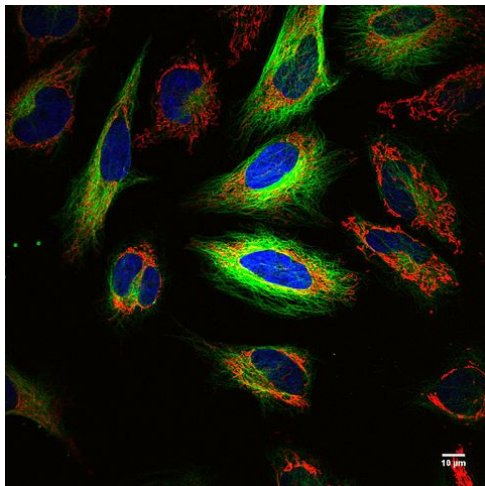
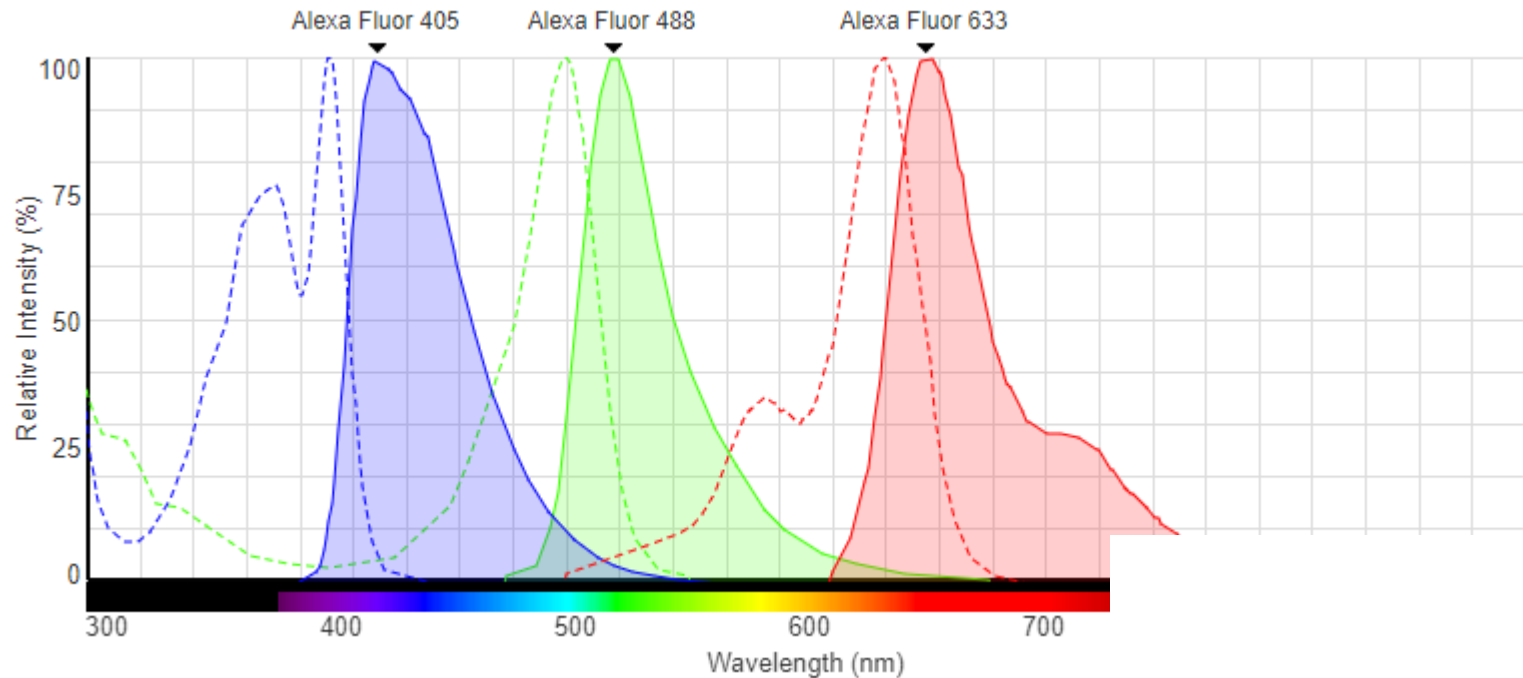
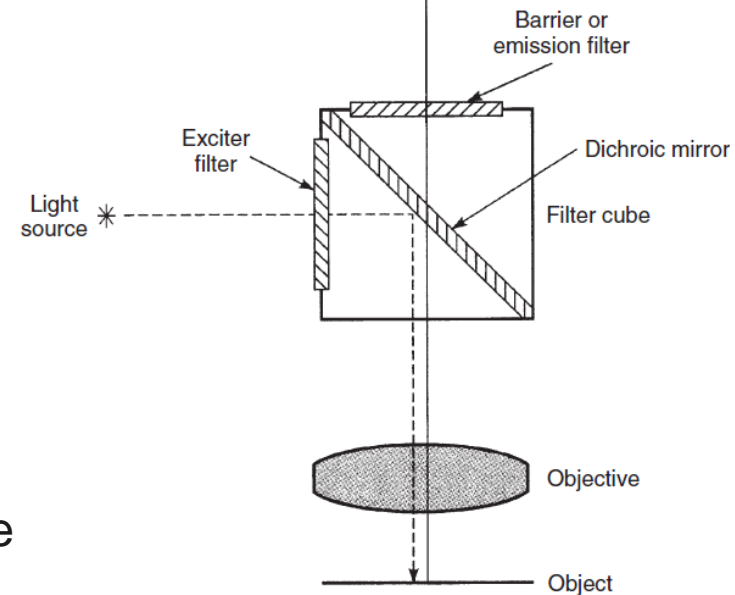


Figure 1

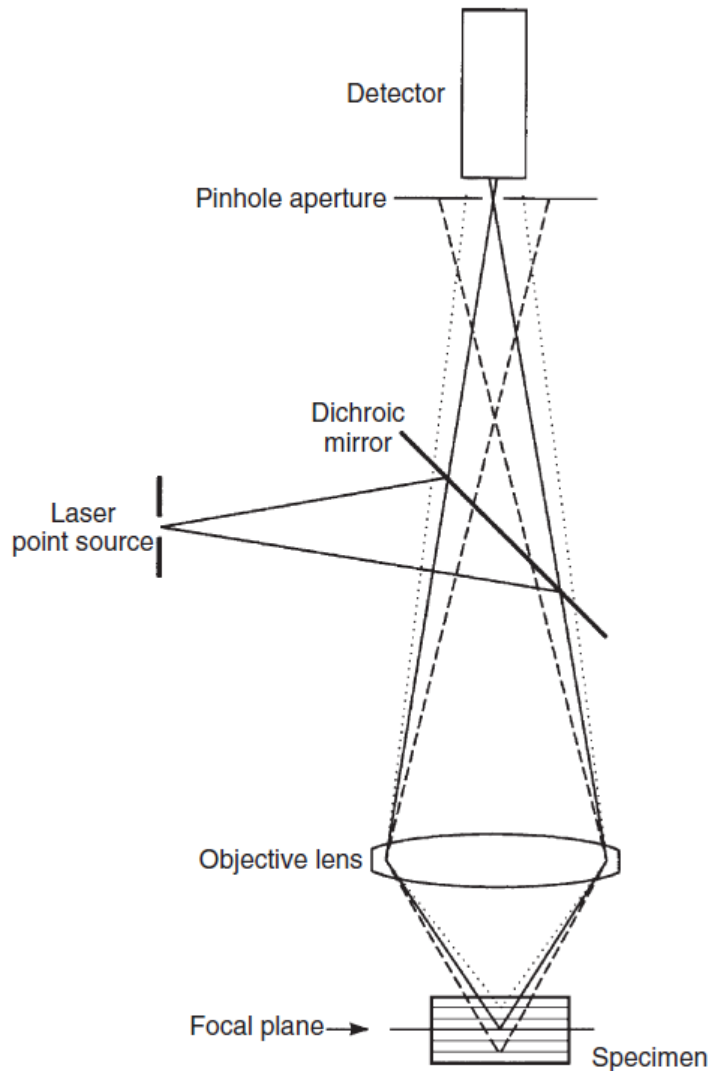
Fluorescence Microscopy



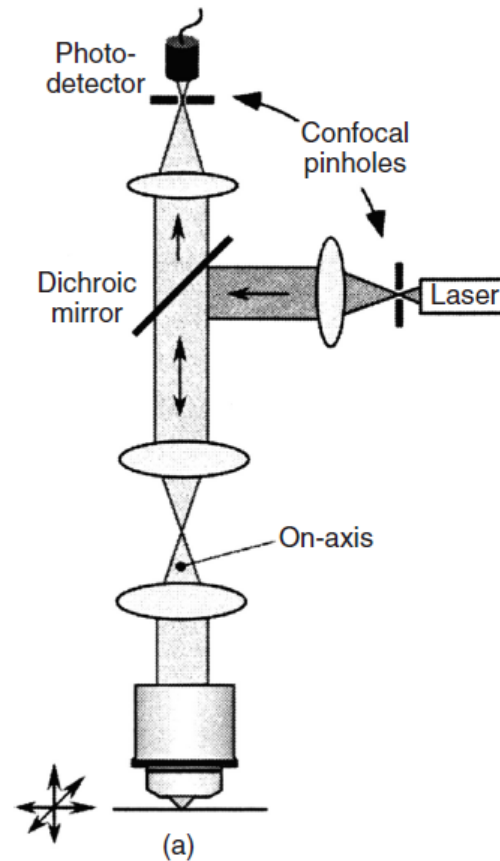
Multicolor fluorescence image of living HeLa cells



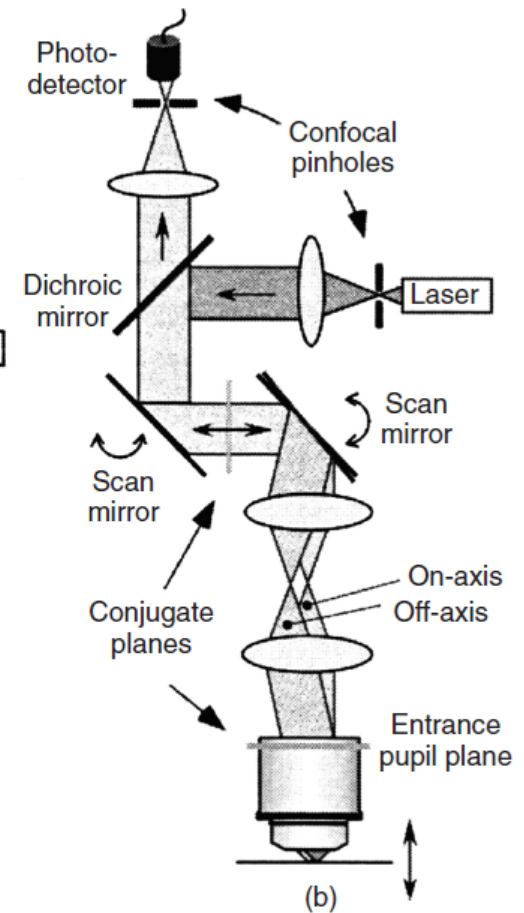
Confocal Microscopy



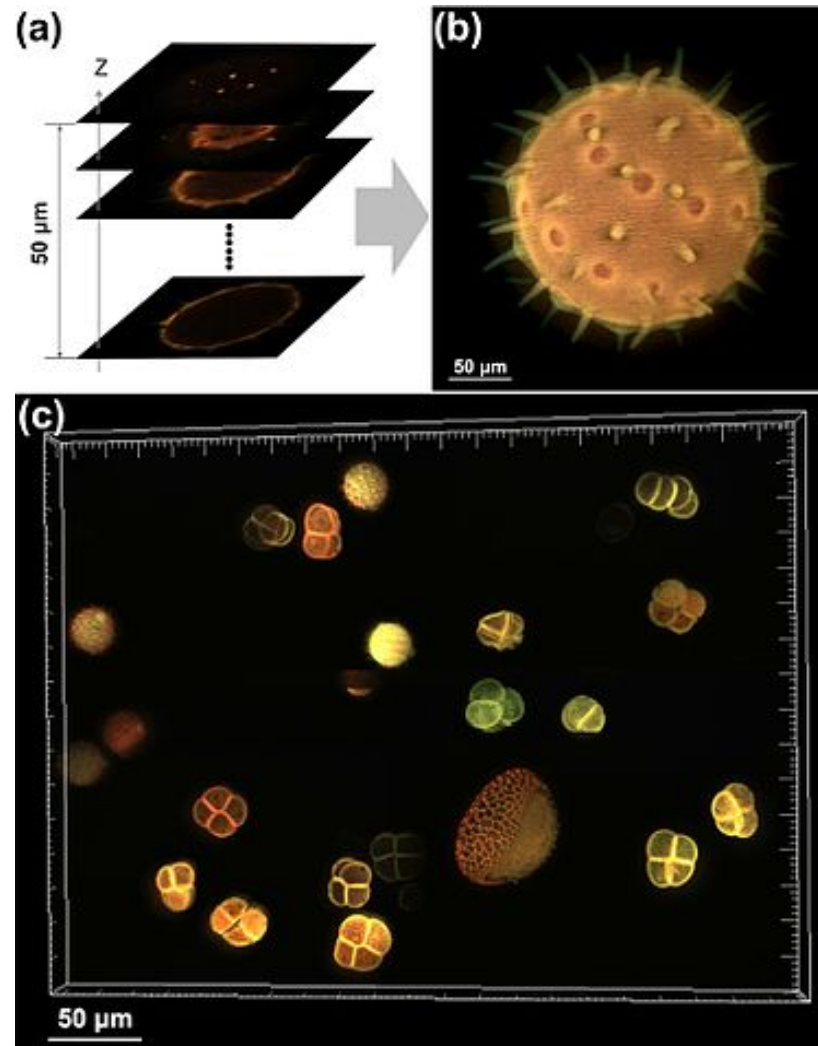
Specimen scanning



Laser scanning



Optical sectioning & 3D image

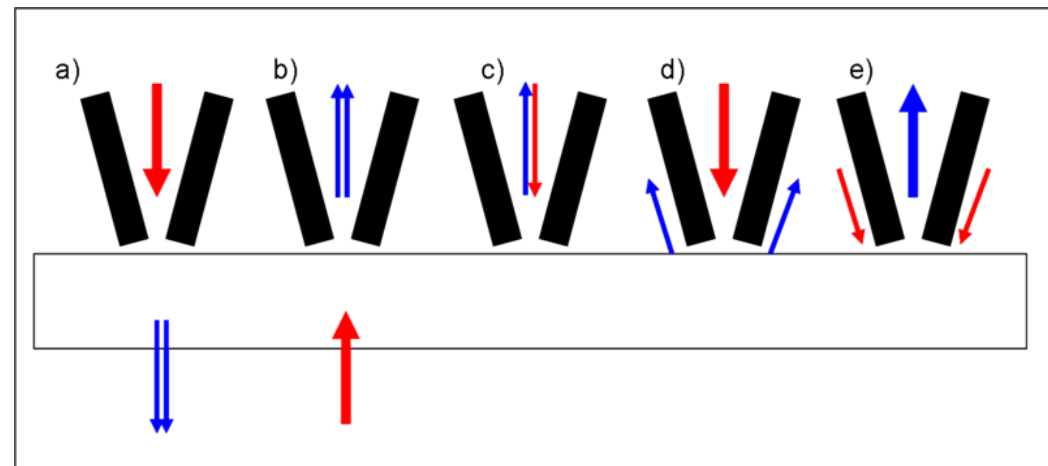
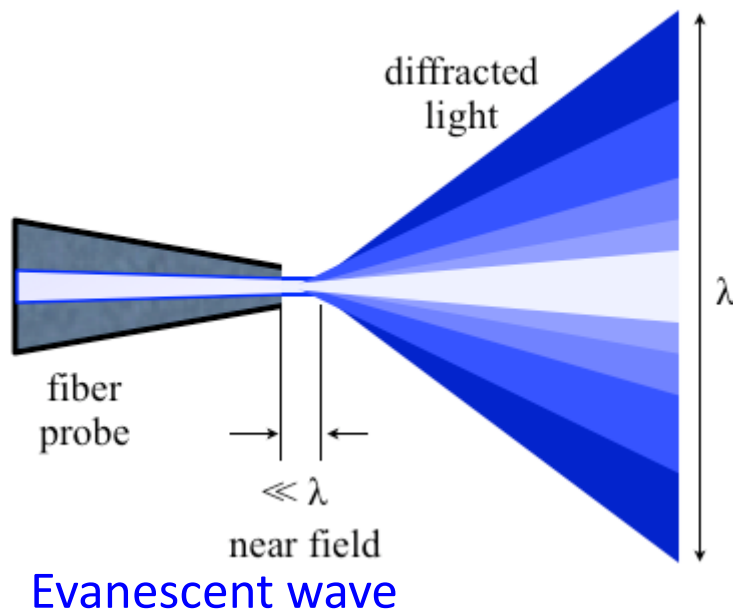


Optical sectioning & 3D fluorescence images of pollens

Optical images at nanometer scale

Break the Abbe diffraction limit ~ 200 nm

- Near-field scanning optical microscopy (NSOM), which is a form of scanning probe microscopy (SPM)



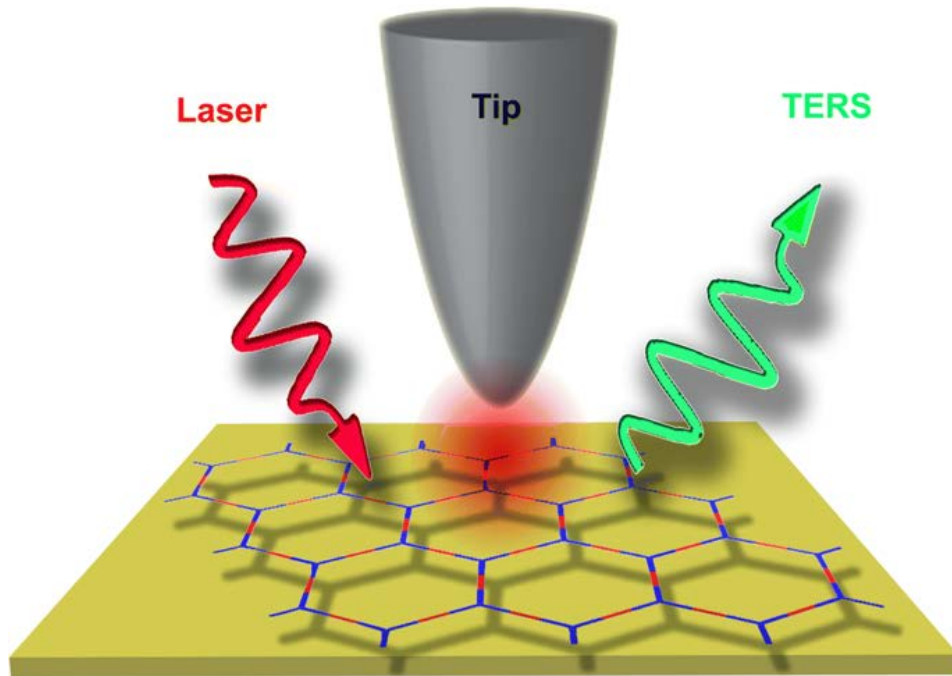
Apertured modes of operation: a) illumination, b) collection, c) illumination collection, d) reflection and e) reflection collection

Optical images at nanometer scale

Break the Abbe diffraction limit ~ 200 nm

- **Apertureless mode:**

Tip-enhanced Raman Scattering (TERS)



Plasmon Resonance:
Enhanced light field!!

Typical enhancement of
Raman Signal: $>10^4$

Far-field optical microscopy with nanometer scale resolution



© Nobel Media AB. Photo: A. Mahmoud

Eric Betzig

Prize share: 1/3



© Nobel Media AB. Photo: A. Mahmoud

Stefan W. Hell

Prize share: 1/3



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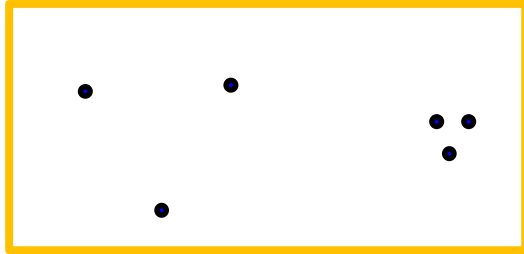
William E. Moerner

Prize share: 1/3

- The Nobel Prize in Chemistry 2014 was awarded for **super-resolved fluorescence microscopy**.

Localization Microscopy

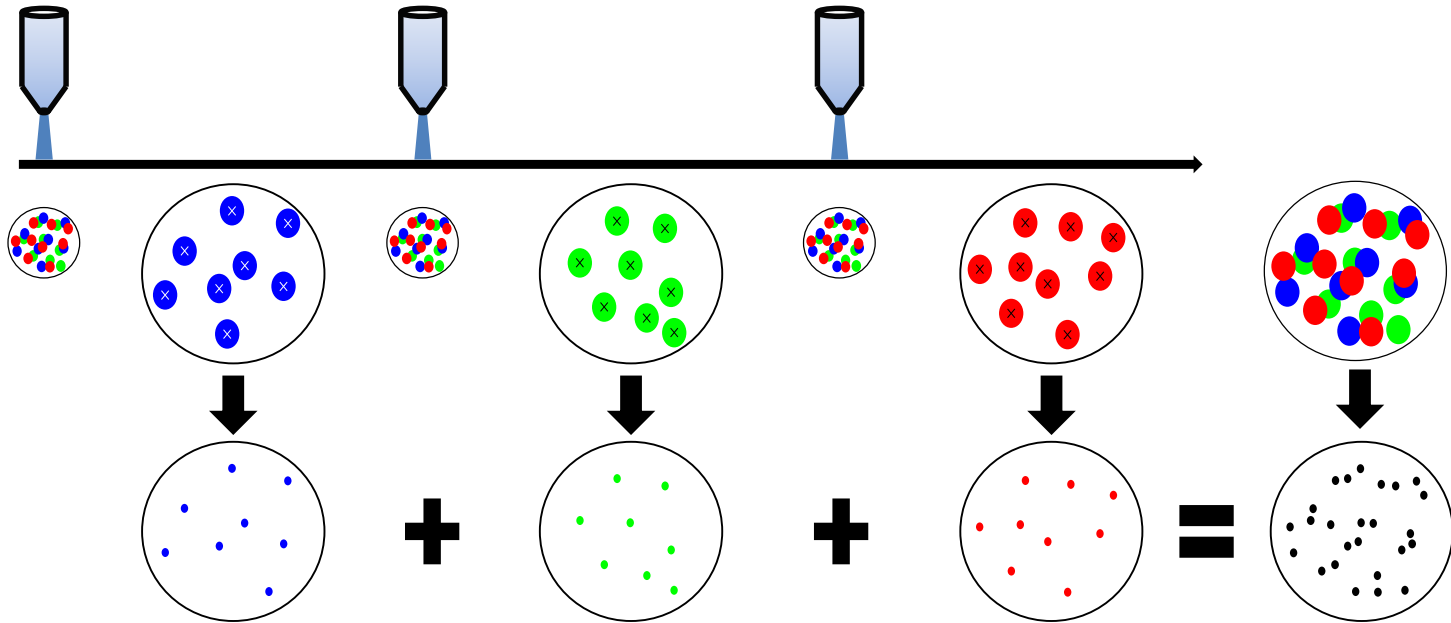
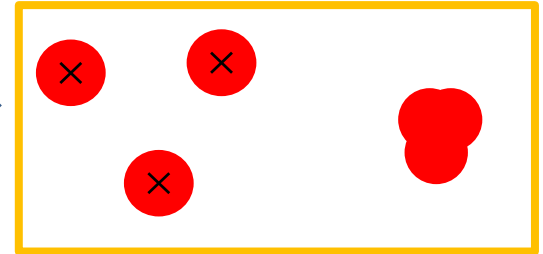
Distribution of molecular dyes



Optical System



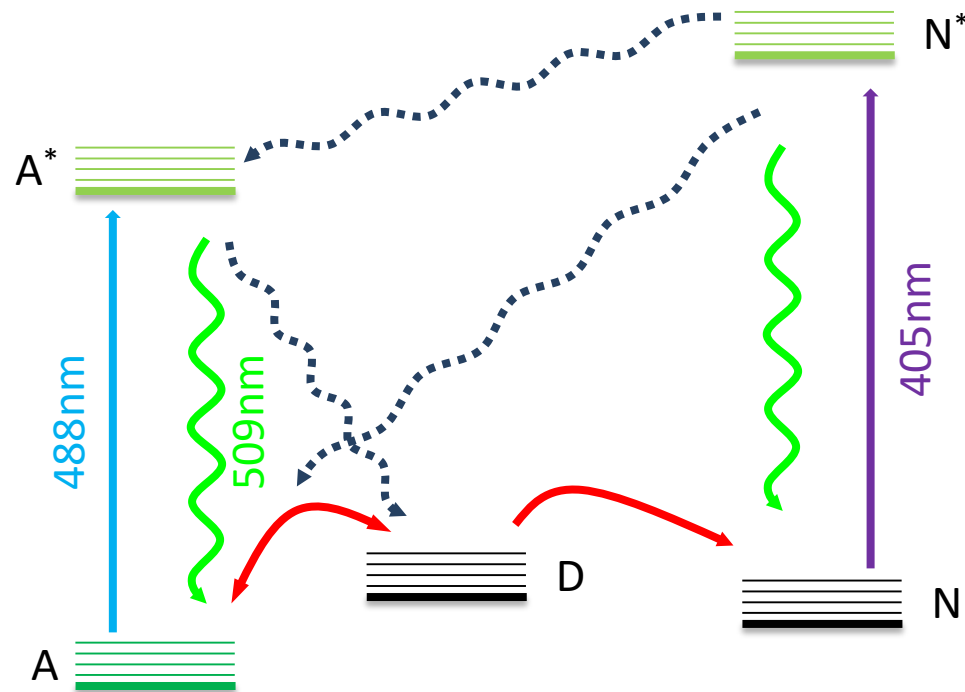
Point spread function (PSF)



**Betzig E (1995) Proposed method for molecular optical imaging.
*Opt Lett.*20:237-239.**

Switching ON/OFF the fluorescence

Green fluorescent protein



**Dickson RM, Cubitt AB, Tsien RY and Moerner WE (1997)
On/off blinking and switching behaviour of single molecules of
green fluorescent protein. *Nature* 388:355-358.**

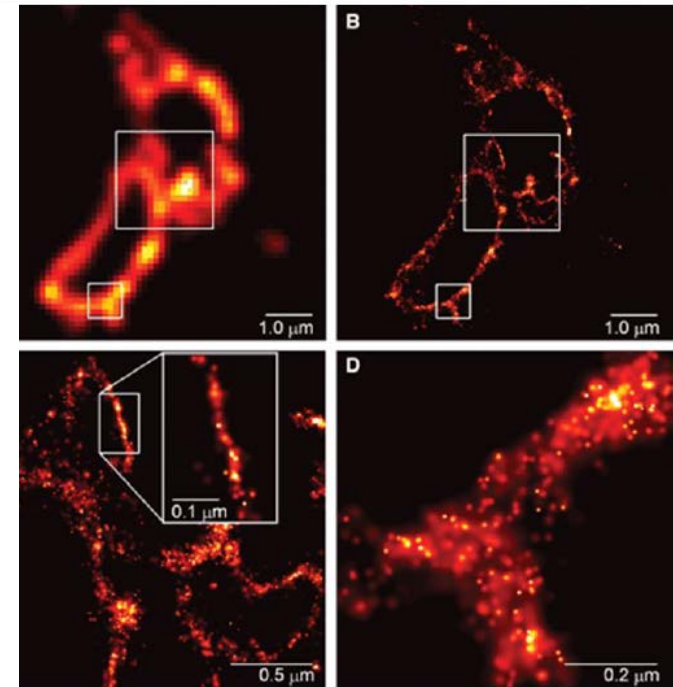
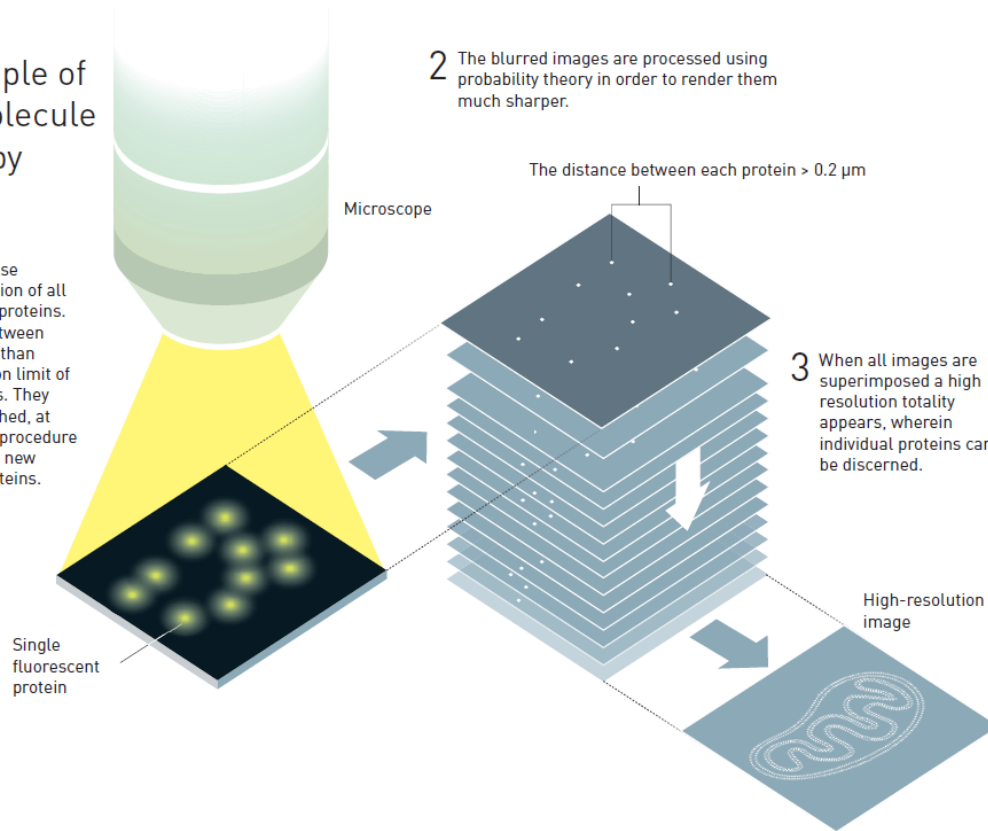
Photoactivated Localization Microscopy (PALM)

Betzig E, Hess HF et al. (2006) Imaging intracellular fluorescent proteins at nanometer resolution. *Science* 313:1642-1645.

Figure 4

The principle of single-molecule microscopy

1 A weak light pulse activates a fraction of all the fluorescent proteins. The distance between them is greater than Abbe's diffraction limit of 0.2 micrometres. They glow until bleached, at which point the procedure is repeated on a new subgroup of proteins.

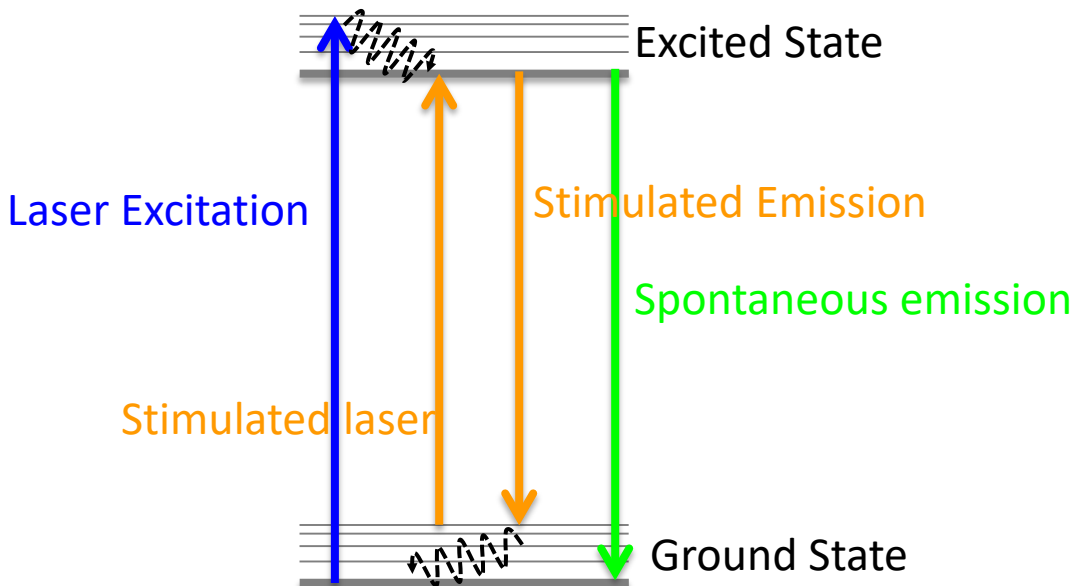


Similar technique: Stochastic optical reconstruction microscopy (STORM)

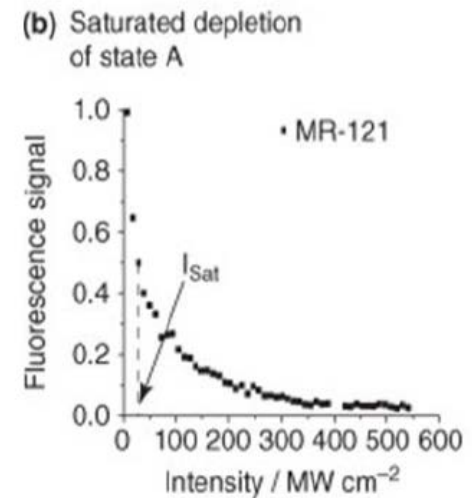
Rust MJ, Bates M and Zhuang X (2006) Sub-diffraction-limit imaging by stochastic optical reconstruction microscopy. (STORM) *Nat Methods* 3:793-795

Switching ON/OFF the fluorescence by Stimulated Emission

Electron energy relaxation

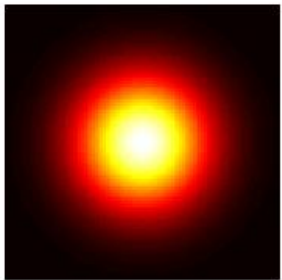


Stimulated emission depletion (STED)

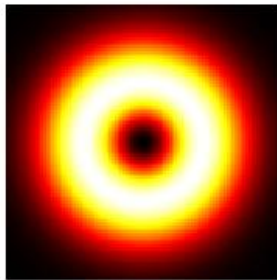


Stimulated Emission Depletion (STED) Microscopy

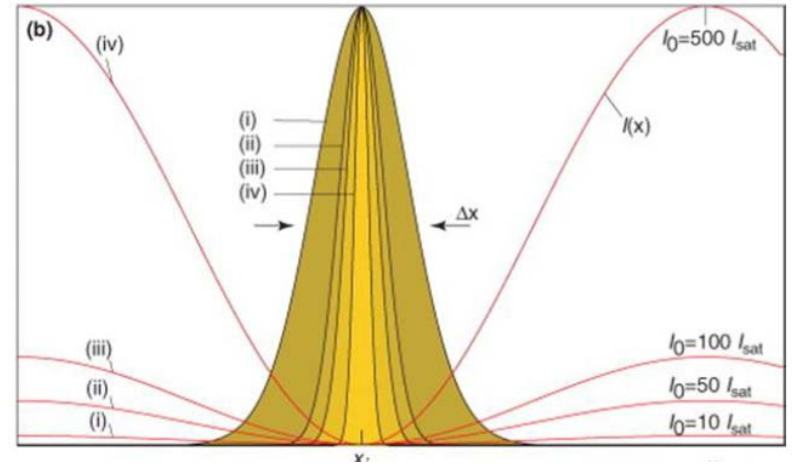
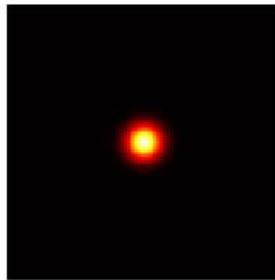
Excitation
Laser spot



STED
laser spot



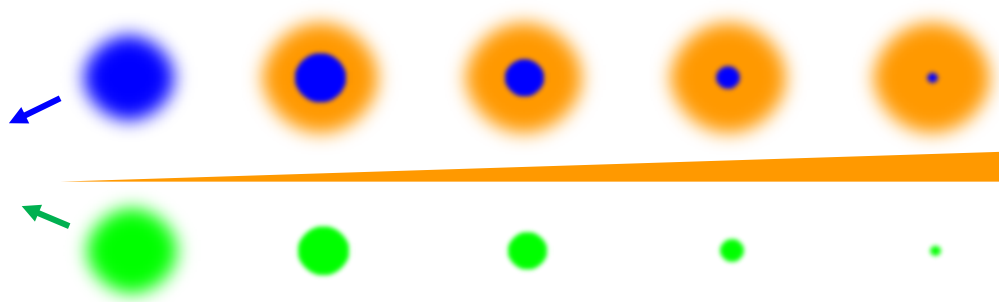
Fluorescence
spot



Excitation

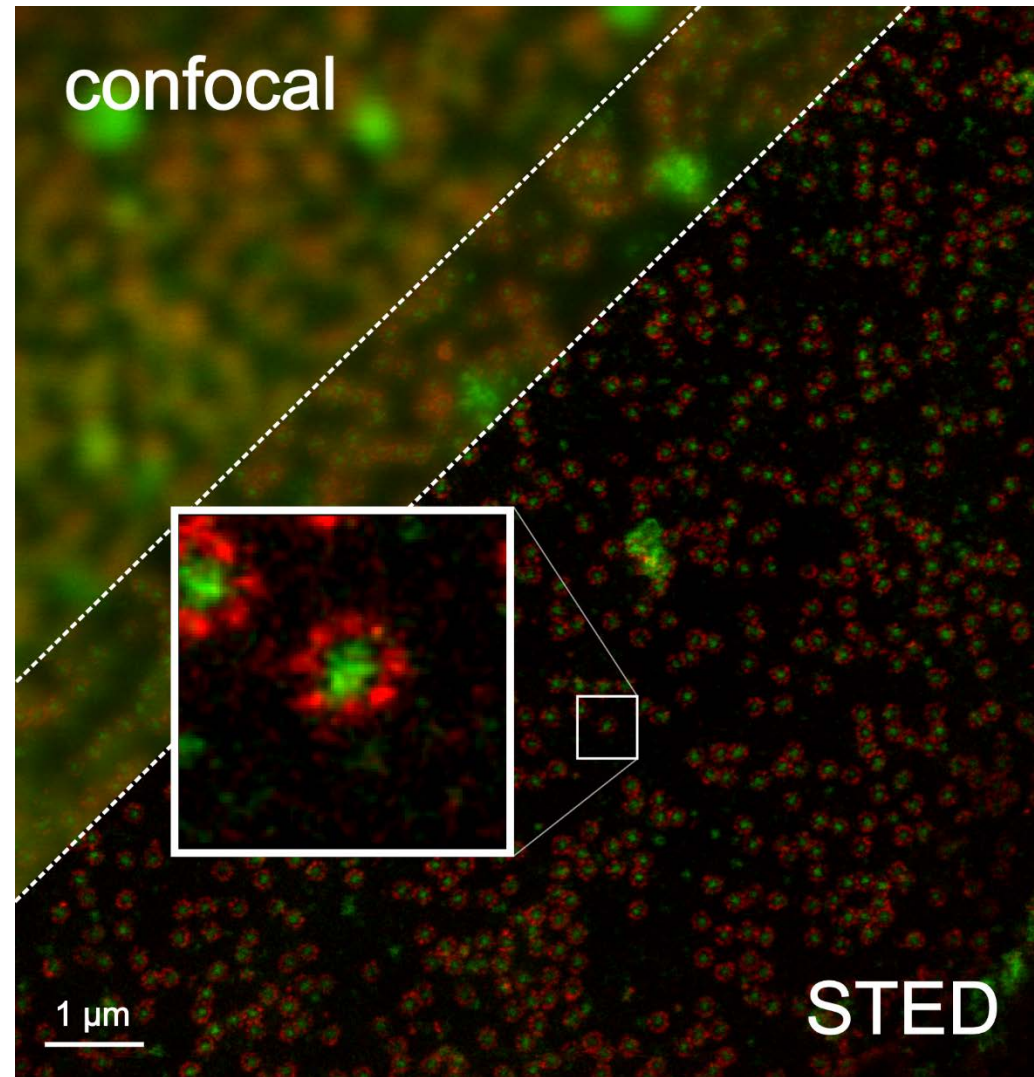
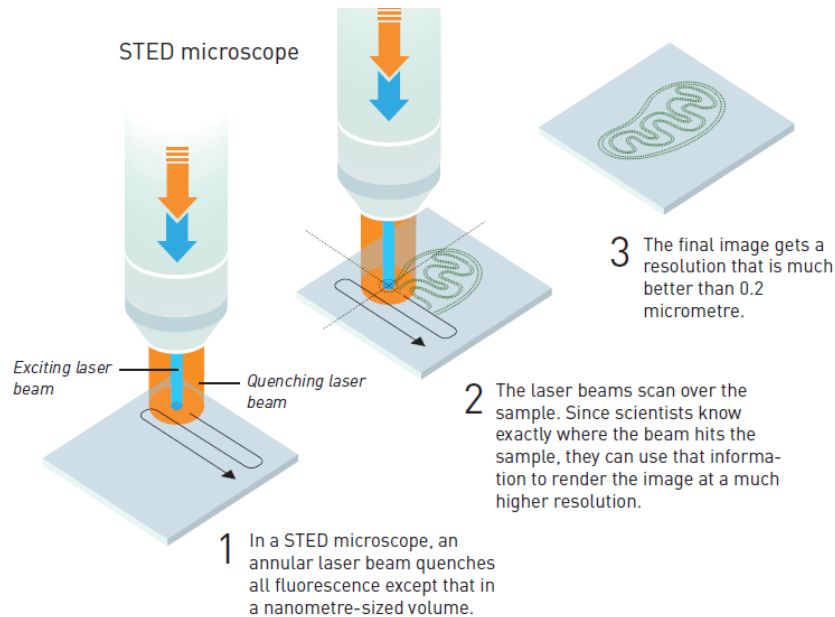
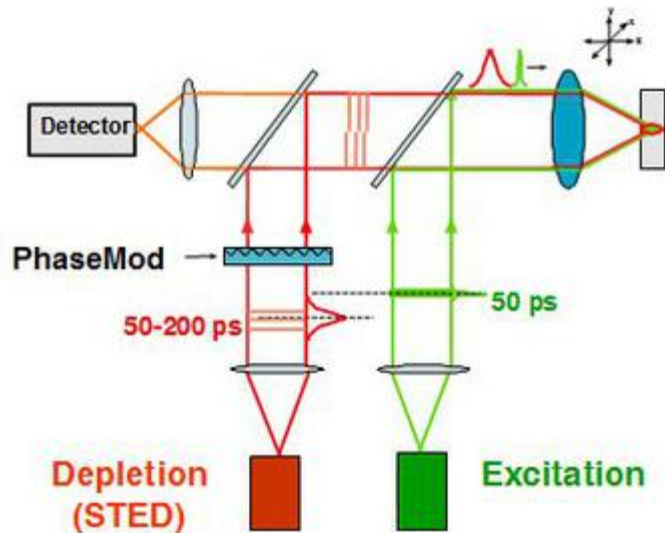
STED spots with donut shape

Diffraction limited
spot sizes



Intensity of STED
Fluorescence

STED Microscopy



Proteins of the nucleus, labelled with fluorescent dyes. **Red: gp210 glycoprotein.**
Green: Several proteins in the central channel

References

- Yang Leng, Materials Characterization: Introduction to Microscopic and Spectroscopic Methods, 2nd Edition, Wiley, Chapter 1
- Olympus Microscopy Resource Center Website
(<https://www.olympus-lifescience.com/en/microscope-resource/>)
- 2014 Chemistry Nobel Prize Website
(<https://www.nobelprize.org/prizes/chemistry/2014/popular-information/>)
(<https://www.nobelprize.org/prizes/chemistry/2014/advanced-information/>)